

# Standard Operating Procedure

## SC22-29 SAMPLING CETACEANS USING A REMOTE BIOPSY SYSTEM

Animal welfare is the responsibility of all personnel involved in the care and use of animals for scientific purposes.

Personnel involved in an Animal Ethics Committee approved project should read and understand their obligations under the *Australian code for the care and use of animals for scientific purposes*.

Version 1.0

August 2022



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August 2022

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The recommended reference for this publication is:  
Department of Biodiversity, Conservation and Attractions (2022). Standard Operating Procedure SC22-29 *Sampling Cetaceans Using a Remote Biopsy System*, Department of Biodiversity, Conservation and Attractions, Western Australia.

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## Revision History

Version	Date	Details	Author/Reviewer	Approved By	Approval
1.0	24/03/2022	Document created	Dr Holly Raudino, Ellen D’Cruz & Dr Kelly Waples		
1.0	31/05/2022	Minor revision	L. Povh	M. Dziminski	August 2022

### Approvals: Version 1.0

Approved by the DBCA Animal Ethics Committee:

**Dr Martin Dziminski**  
Chair, Animal Ethics Committee  
Department of Biodiversity, Conservation and Attractions

# Contents

1	Acknowledgement.....	3
2	Purpose .....	4
3	Scope.....	4
4	Animal Welfare Considerations.....	5
4.1	Level of Impact.....	5
5	Procedure Outline .....	6
5.1	Equipment, care, and maintenance .....	6
5.2	Personnel .....	10
5.3	Sterilisation and safety procedures.....	10
5.4	Sampling technique.....	12
5.5	Data recording.....	16
6	Competencies and Approvals .....	17
6.1	Occupational Health and Safety .....	18
6.2	Personal Protective Equipment.....	18
6.3	Manual handling and working with ropes .....	19
6.4	Working around the water’s edge and from vessels .....	19
7	Further Reading.....	19
8	References.....	20
9	Glossary of Terms.....	20

# 1 Acknowledgement

Advice for compiling the content of this standard operating procedure has been generously provided over the years from Delphine Chabanne, Anna Kopps, Alex Brown, Simon Allen, and Daniel Barrow. The procedures have been trialled by DBCA staff across the Pilbara and Kimberley areas, including participation of joint managers and traditional owners. We would like to acknowledge the traditional owner's past, present and emerging of the land and sea country we work on, Dambimangari, Murujuga, Yawuru, and it is an honour to work with these groups on their sea country. Thanks to DBCA staff and vessel crew throughout the Kimberley and Pilbara regions that have contributed to successful and safe surveys over the years.

## 2 Purpose

Tissue samples collected from cetaceans can be used to investigate biological and ecological questions about the animals. DNA can be extracted from these samples and used to identify species, phylogeography, gene flow and population structure through genetic analyses. Food web analyses and dietary studies can also be undertaken using stable isotope and fatty acid signature analyses and environmental and anthropogenic pollutants can be explored with contaminant/biomarker analyses. Collecting tissue samples from live, free-ranging animals can be invasive and often requires the animal to be captured. However, there are methods available that allow for remote sample collection. This Standard Operating Procedure (SOP) describes a remote sampling technique used to collect tissue from free-ranging cetaceans using a biopsy rifle and dart or biopsy pole. While the equipment and its use are detailed within this document, it is not intended to be a stand-alone reference for cetacean work and would typically take place during other field work such as vessel-based cetacean surveys (*SOP: Vessel-based Cetacean Surveys Using Photo Identification*).

Much of this information is derived from techniques that are commonly used for this type of research and on the experience of research scientists in the Marine Science Program supplemented by work by regional staff, Murdoch University and Alex Brown.

## 3 Scope

This SOP has been written specifically for scientific and education purposes and endorsed by the department's Animal Ethics Committee. However, this SOP may also be appropriate for other situations.

This SOP applies to all remote biopsy sampling of cetaceans undertaken across the State by Department of Biodiversity, Conservation and Attractions (hereafter department) personnel. It may also be used to guide fauna monitoring activities undertaken by Natural Resource Management groups, consultants, researchers and any other individuals or organisations. All department personnel involved in remote biopsy sampling of cetaceans should be familiar with the content of this document.

Projects involving wildlife may require a licence under the provisions of the *Biodiversity Conservation Act 2016*. Personnel should consult the department's Wildlife Licensing Section and Animal Ethics Committee Executive Officer for further guidance. In Western Australia, any person using animals for scientific purposes must also be covered by a licence issued under the provisions of the *Animal Welfare Act 2002*, which is administered by the Department of Primary Industries and Regional Development. This SOP complements the *Australian code of practice for the care and use of animals for scientific purposes* (The Code). The Code provides governing principles to guide decisions and actions of personnel involved in the care and use of animals and contains an introduction to the ethical use of animals in wildlife studies. A copy of The Code may be viewed by visiting the National Health and Medical Research Council website (<http://www.nhmrc.gov.au>).

## 4 Animal Welfare Considerations

To reduce the level of impact on the welfare of animals, staff must consider, address and plan for the range of welfare impacts that may be encountered. Strategies to reduce impacts should be identified during the planning stage to ensure that they can be readily implemented during activities and contingencies for managing welfare issues have been identified. All personnel involved in the project should be aware of the range of issues that they may encounter, the options that are available for reducing impacts and improving animal welfare, and the process for managing adverse events.

Department projects involving cetacean biopsy sampling will require approval from the department's Animal Ethics Committee.

### 4.1 Level of Impact

#### ***Environmental***

There are two main ways in which an animal may be impacted by the biopsy procedure: they may be disturbed by the actual sampling, or they may be harmed by the wound created by the dart, or they may be disturbed (react) to the actual sampling or the close approach of the vessel.

Darting wounds typically heal rapidly and without complication (Tezanos-Pinto & Baker 2012; Weller et al., 1997). Photographs collected during sampling events should be used to identify the individuals sampled and to, where possible, confirm the sampling site on the animal at time of sampling. These can then be used to ensure normal healing of the biopsy wound is occurring if the animal is re-sighted in days, months and years after.

There are different stages to the biopsy wound healing process that can be monitored if repeated photographs are available. In the first stage the wound is covered, smooth and absent of pink colouration which generally occurs within weeks to up to 3 months (as a maximum time frame) (Weller et al., 1997). For example, Krutzen et al. (2002) found wounds were healed after 23 days in individual bottlenose dolphins that they observed daily post-biopsying. Re-pigmentation takes a lot longer, months and in some cases years in other delphinids such as pilot whales (Gimenez et al., 2011). Another study measured 10 individuals from 3 days to 7 months post biopsying and healing appeared to occur without complication; no bleeding, swelling or signs of infection were observed (Tezanos-Pinto & Baker 2012). Based on the collective literature we are expecting that healing will vary between individuals but will range between two weeks and three months for the epidermis to cover the small wound created by the biopsy dart which is superficial compared to large deep wounds from shark encounters that also heal relatively quickly (Corkeron et al., 1987; Orams & Deakin, 1997).

Response to the procedure will be monitored. Darting reactions are generally mild and short-term ('flinch' or 'buck'), and most animals continue their pre-biopsy behaviours within the area (Krutzen et al., 2002) an extreme aversive reaction would be a leap. There has only ever been one mortality from a remote biopsy attempt documented in the literature and this was thought to potentially be the result of vertebral trauma due to thin blubber layer ~7mm (poor body condition) and/or stress (Bearzi, 2000). In the event of an unexpected death, the carcass will need to be retained for necropsy and the Animal Ethics Committee notified within 24 hours and a comprehensive report written on the adverse event following the initial

notification. The biopsy 'procedure' is instantaneous and ideally occurs within the first 15 minutes of encountering the group. Sampling should be ceased if any animals show active avoidance of the vessel. Further, no more than three attempts will be made per individual.

Exposure to vessels and the associated motor noise can disturb cetaceans and disrupt critical activities such as feeding, resting and nursing and attending to calves. Vessels should be handled in a way that minimises disturbance to the targeted fauna. Time limits will be set and complied with. The research vessel will spend no more than 60 minutes within 100m of a cetacean group, regardless of whether all group members have been photographed and/ or biopsied. Time limits and approach distances are prescribed under the Biodiversity Conservation Regulations 2018 and further licence conditions may be in scientific licences. Other considerations may be around the presence of young e.g., newborn calves or in critical areas or at critical times i.e., calving and nursing. The duration of encounters should be further limited if groups contain newborns or groups are resting or nursing young calves to prevent disruption of critical activities.

### ***Social***

There may be live-aboard vessels or other commercial operations occurring within the survey area. It is recommended that effort is made to contact or approach these vessels prior to conducting the survey to explain the operation, and if required determine how to work around any commercial operations such as barramundi netting, crabbing, pearling, tourism interactions etc. If commercial or recreational vessels enter the survey area during a survey, particularly if biopsying is occurring, they should be contacted via radio communication to explain the on-going operations and told not to approach animal groups closer than 100 metres while DBCA vessels are photographing, biopsying and observing.

## **5 Procedure Outline**

### **5.1 Equipment, care, and maintenance**

#### ***Biopsy rifle***

There are two rifle systems that have been used by DBCA researchers to biopsy cetaceans; the Dan-Inject and the PAXARMS. Both systems are described below.

#### ***PAXARMS***

This system is a modified 0.22 calibre rifle with Pro-Pointa red-dot sight and uses blank charges to propel a lightweight, polycarbonate plastic biopsy dart at free-ranging cetaceans. It consists of a stock, detachable barrel, and a pressure (distance-) adjustable valve system (Figure 1a) that propels a biopsy dart (Figure 1b). This system (PAXARMS: <http://www.paxarms.co.nz/>) is well established and has been used extensively internationally (e.g., Krützen et al., 2002) as well as around Australia for a number of years.





Figure 1 The PAXARMS biopsy system. (a) Assembled 0.22 caliber rifle with Pro-Point red-dot sight. (b) Biopsy dart and biopsy tip. Photos taken from Murdoch University Cetacean Research Unit.

The PAXARMS darts have a hollow body with a thin, red polycarbonate plastic tube exterior and steel biopsy tips bevelled inwards (Figure 1b). Three small triangular barbs are located inside the steel tip 2 mm from the leading edge of the tip, for sample retention. A flat metal rim (flange) is welded to the base of the biopsy tip directly above where it screws into the body of the dart. The flange acts as a stop to ensure the dart does not penetrate too far into the animal. The body of the dart is a hollow plastic tube and there is an internal wall slightly below the thread for the flange to prevent flooding of the tube. A small piece of polycarbonate plastic screws into another thread at the tail of the body that acts as a safety partition if the tail separates on impact (Figure 2a). The polycarbonate plastic tailpiece has an o-ring and screws into the tail end after the partition to create a watertight seal.

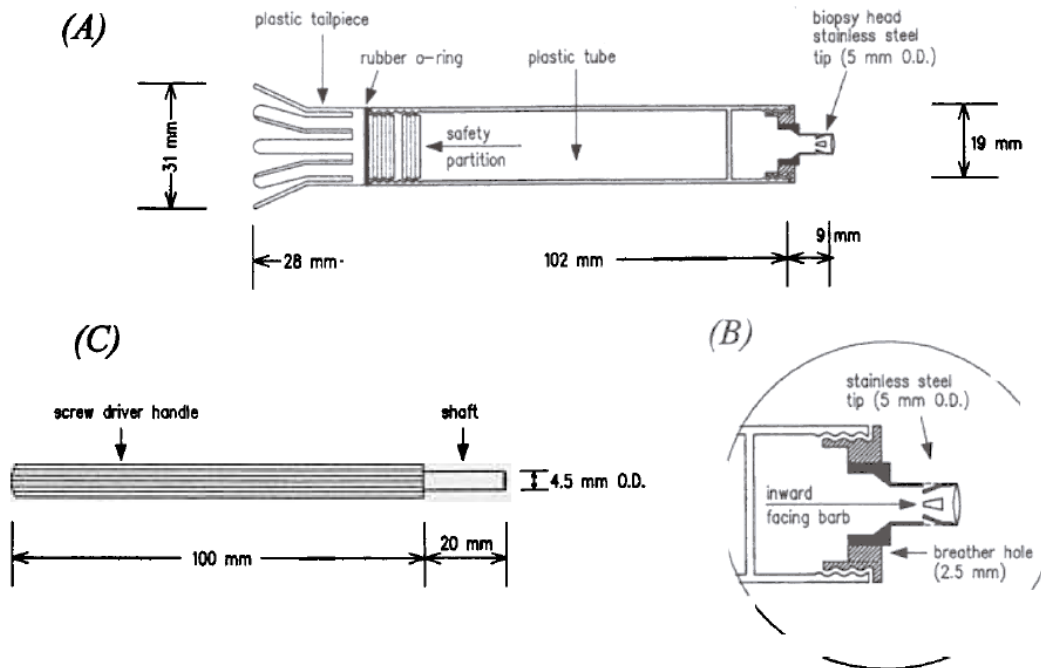


Figure 2 Biopsy dart (A) Assembled biopsy dart showing inserts of biopsy tip, safety partition, and plastic tail. (B) Detailed drawing of biopsy tip. (C) Barb resetting tool. O.D. = outer diameter (taken from Krützen et al., 2002).

The biopsy darts are positively buoyant, float in an upright position and have a total assembled weight of approximately 21.5 g. Blank charges are used to project the darts and proprietary charges can be purchased from PAXARMS or alternatively some brands of nail gun cartridges are suitable (see Ramset Australia Power loads (5.6 x 16mm calibre) medium (PLCYW22S) or strong (PLCRD22S) single cartridges). A dial on the side of the rifle chamber is used to adjust the pressure of the dart projection. A red-dot laser sight (Tasco) is fitted to the rifle to improve darting accuracy, as darting is faster and more accurate with this system than an open sight. A barb-setting tool (Figure 2c) should be used to reset the barbs. With an unscrewed steel biopsy tip, the tool should be inserted from the underside and pushed forward to reset the barbs regularly.

#### Dan-Inject

This system is a Carbon Dioxide (CO<sub>2</sub>) gas powered rifle with a red laser sight and uses CO<sub>2</sub> to propel the plastic syringe with biopsy dart head at a free-ranging animal (Figure 3). The Dan-Inject (<https://dan-inject.com/> or <https://daninject.com.au/>) has been used for skin tissue sampling on a variety of free-ranging animals (Mijele et al., 2016). The 5mm diameter off-the-shelf dart heads were customised for use with cetaceans by reducing the shaft length of the dart head to 8mm and adding a metal washer / stopper below this to prevent the dart from embedding into the animal

The darts have a hollow body with a white transparent plastic tube exterior and steel biopsy bevelled tip (Figure 3). The hollow rear chamber of the dart body (plastic syringe) causes the dart to float tail-end up and the bright pink synthetic tail makes them relatively easy to find. Two triangular barbs are located slightly in from the tip edge for sample retention. The system has a built-in CO<sub>2</sub> pressure gauge oriented on the rifle facing the shooter and a pressure

control valve that can be monitored and adjusted while aiming. Forceps or a probe tool (or similar) can be used to push the barbs inward to reset them regularly.

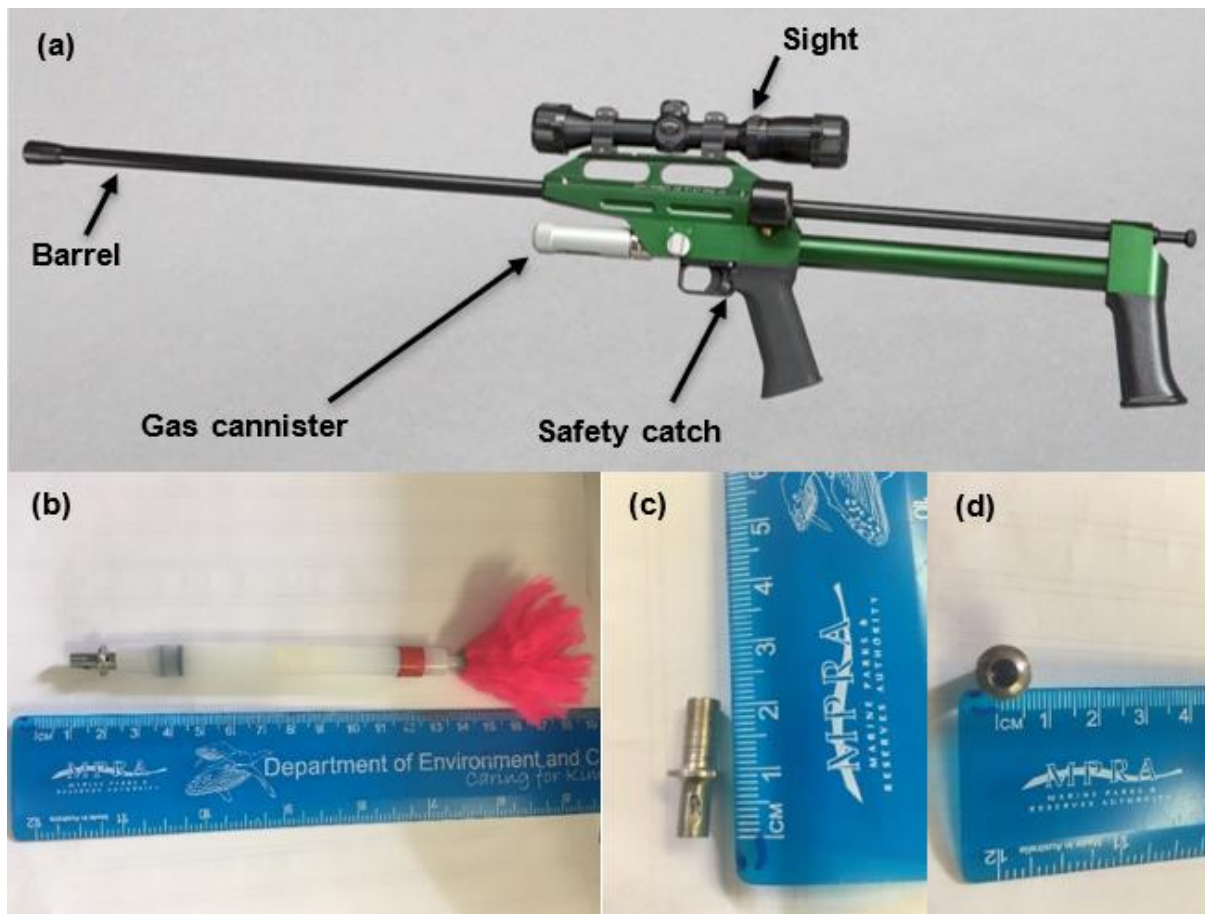


Figure 3 Dan-Inject biopsy system. (a) Assembled rifle with Pro-Point red-dot sight. (b) dart head with syringe and tail, (c) prongs and (d) slits added to the dart head in 2019 to improve tissue sample retention.

DBCA personnel (with appropriate firearms training, competency and currency as a nominated person on the DBCA corporate firearms licence for Category E firearms) have used both the PAXARMS and Dan-Inject system to sample cetaceans across the State. The PAXARMS is tried and tested and has been successfully used to obtain tissue samples from several dolphin species, whereas the Dan-inject dart heads require further modification to make them suitable for sampling small cetaceans (dolphins) successfully. Both systems explained above are hereafter referred to as the biopsy rifle.

#### *Biopsy pole*

The pole head is similar to the biopsy dart head previously described and similarly is used to take a small plug of skin and blubber (Bilgmann et al., 2007). The dimensions of the biopsy pole are head 30 mm width; biopsy tip 6mm width with bevelled edge (Figure 4 taken from Bilgmann et al., 2007). The biopsy tip is a hollow cylinder that has barbs to hold the sample in place. Water that flows into the pole through the tip flushes out through a purge hole further up the pole. This prevents it from flowing back down the shaft and dislodging the sample.

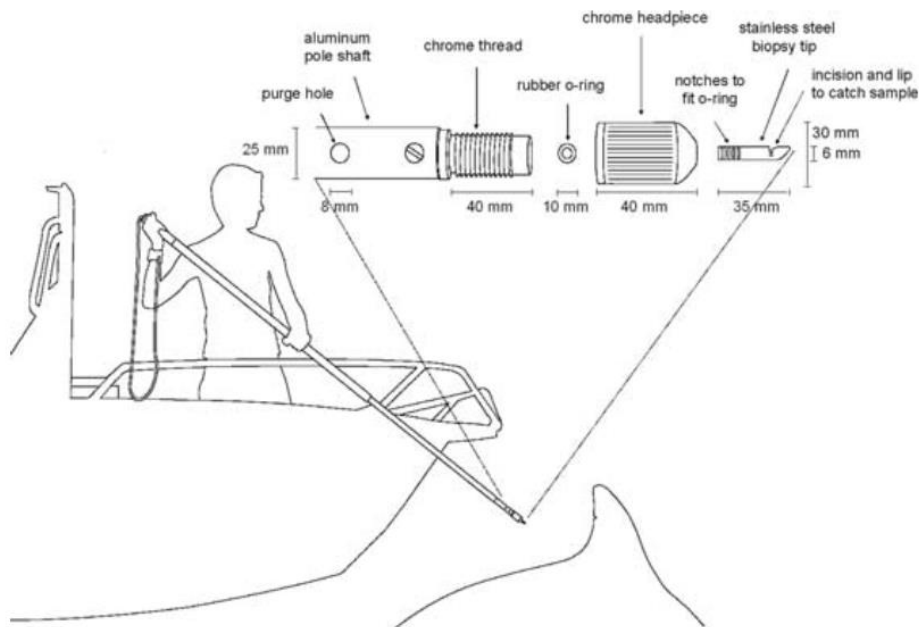


Figure 4 Lateral schematic of technique and detail of distal components of the biopsy pole system including biopsy tip (taken from Bilgmann et al., 2007)

## 5.2 Personnel

There are three essential personnel required for remote biopsy of cetaceans using either the rifle or the pole system:

- The shooter stands in the bow of the vessel, ready to biopsy
- The observer/photographer stands behind the shooter taking identification photos of the animal at the same time as the individual is being sampled (so that it can retrospectively be identified)
- The vessel master is responsible for safety and navigation.

A second observer (optional) is also valuable as they can be responsible for recovering darts using a hand-net, storing samples, recording notes, and acting as a secondary photographer (if two cameras are available).

## 5.3 Sterilisation and safety procedures

The following sterilisation procedure should be completed before starting the survey each day. This procedure **MUST** take place away from fuel and other flammables and close to a water supply and a suitable fire extinguisher (i.e., CO<sub>2</sub>):

1. If facilities are available in the field, the biopsy heads should be boiled for 10-20 minutes and then, using forceps, biopsy heads should be submerged in 95% Ethanol and then flamed (lit and extinguished) with a lighter. If boiling is not possible, the biopsy heads should be scrubbed with a clean toothbrush (or similar) and flamed and the biopsy heads allowed to cool. Biopsy heads should never be touched on the biopsy

tip or cutting edge. Biopsy heads should only ever be touched with gloved hands after sterilisation (scrubbed and flamed) around the thread while being screwed onto the dart bodies. The biopsy heads can be easily grasped and manipulated with forceps and manipulated by the dart body once the heads are screwed back onto the dart body. Biopsy heads should be checked and re-sharpened and barbs reset with appropriate tool at time of cleaning process if required.

2. Sterilised biopsy heads should be stored in a clean, watertight container.
3. Sterile latex gloves must be worn when handling biopsy heads at any time, to minimize the risk of infection to both the dolphin and observer and cross-contamination by DNA/RNA from either and other samples. Gloves should be changed between sampling events/animals.

Safe rifle procedures while on board a vessel are outlined below, but specifications may differ between rifle types not covered by this SOP:

1. Place rifle into vessel in a place where it is secure and protected from water spray, wash and in preferably a secure case i.e., lockable pelican case.
2. When a suitable group of cetaceans has been identified the rifle can be removed from the case and assembled [Note: only a person with the appropriate Firearms Licence and experience with the firearm being used can handle the firearm]:
  - a. Attach barrel to biopsy rifle stock
  - b. Insert dart into barrel
  - c. Load blank cartridges into magazine and insert magazine into rifle (PAXARMS) or screw in CO<sub>2</sub> cannister (Dan-inject).
3. Use biopsy rifle only from the bow of the vessel, ensure crew always remain behind shooter. General rule do not shoot any further back than mid-ship of the vessel or 3 and 9 o'clock on the clock face if the bow is 12 o'clock and the stern is at 6 o'clock (Figure 5).
4. Before darting, cock the rifle, and ensure safety catch is on.
5. The shooter must always communicate with the crew to ensure that all personnel are aware that the dart is about to be fired. Once the shooter has determined that a sampling attempt is appropriate and the cetacean group has been approached, release the safety catch and line up the sight on the lateral side of the animal, below the dorsal fin (see Krützen et al., 2002). Only fire when safe to do so for both the target animal, and people in the vessel.
6. After a darting attempt, place rifle in a safe, dry place and recover biopsy dart using a hand-net (ideally this is by someone other than the shooter).
7. Remove sample from dart taking care not to touch the sample with anything other than sterilized tools and place in an individual sample vial. If the sample is stuck, the biopsy head may need to be removed and the sample carefully pulled out using forceps from the back of the biopsy head. Sterile tooth picks (or piksters) or a stainless-steel probe may be useful for extracting the sample.
8. Label the sample vial clearly and store sample away from heat (if possible, in a cooler box for samples) and complete the associated biopsy datasheet.
9. Once equipment has been used (i.e., forceps, probes, biopsy heads) to collect a sample these need to be stored separately from the remaining sterilised equipment.

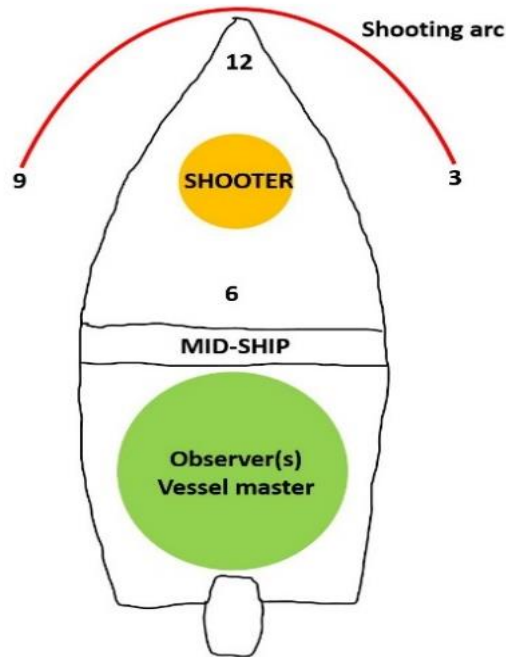


Figure 5 Overhead schematic of how personnel should be positioned on the survey vessel and the shooting arc

10. Label the sample vial clearly and store sample away from heat (if possible, in a cooler box for samples) and complete the associated biopsy datasheet.
11. Once equipment has been used (i.e., forceps, probes, biopsy heads) to collect a sample these need to be stored separately from the remaining sterilised equipment.
12. Between sampling events ensure there is no dart in the rifle chamber.
13. If sampling more than one animal in a group, repeat from step 2 with sterilised biopsy dart.
14. When all sampling has been completed with the cetacean group, disassemble the rifle, and return it to its case and a safe dry location on the vessel.
15. Clean and store rifle according to manufacturer's instruction at the end of survey day. Store biopsy rifle in rifle case, ensuring no blank cartridges are left in the rifle. Ammunition and rifles are stored securely (under lock and key when not in use) and separately.

A miss-shot is likely to be dissipated by the water surface. To prevent misfiring in the vessel the following precautions are used (as described above in Section 4.3): safety catch is used, crew stand behind the person using the rifle, the rifle is stored in an appropriate case when not in use. If the dart completely misses an animal, more attempts can be made if the animal shows no signs of behavioural changes (i.e., vessel avoidance or other reaction) and can still be approached. Further, if any animals in the group (incl. sampled individuals) show signs of behavioural changes indicating disturbance (e.g., repeated rapid surfacing and dives in a direction away from the boat), sampling should be ceased and the vessel move away (Krutzen et al., 2002).

#### 5.4 Sampling technique

Biopsy sampling can only be conducted in suitable sea conditions (no rain, light winds, and seas less than 1.0m i.e., equivalent of Beaufort Sea State [BSS]  $\leq 3$ , Table 1) and should be undertaken from small vessels (i.e., <7m). Speeds should be reduced to no wake or neutral particularly when close to groups (e.g., within 50m). The animals are aware of the vessel when the motor is running, so it should not be turned on and off when interacting with the group. Sampling should only be attempted when animals are between approximately five and 20 m from the vessel and the pressure adjusted on the rifle accordingly. Photographs of the dorsal fin of the animal(s) to be sampled will be taken before and during sampling for the purpose of photo-identification of individual animals and to match the sample with that individual. For some large whale species, other features can be used for identification and should be photographed. This may include underside of flukes for humpback whales, the callosity pattern on the head for southern right whales, and the lateral pigmentation pattern for blue whales. Refer to SOP No: TBA Vessel-based Cetacean Surveys Using Photo Identification for detailed instructions on photographing cetaceans and data collection.

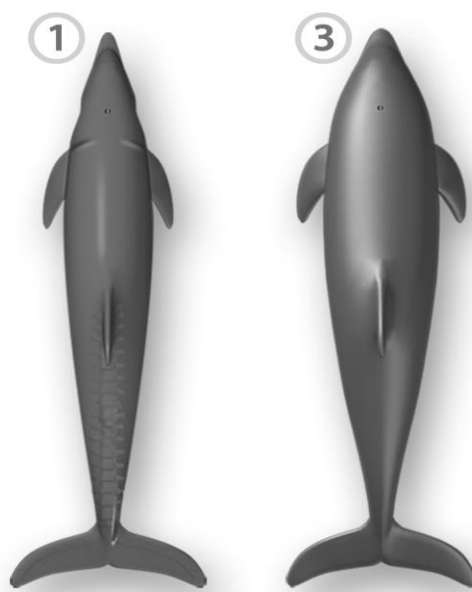
Table 1 Beaufort Sea State scale description.

Beaufort number	Wind speed (knots)	Wave height (m)	Sea conditions
0	< 1	0	Flat
1	1-3	0 - 0.2	Ripples without crests
2	4-6	0.2 – 0.5	Small wavelets. Crests of glassy appearance, not breaking.
3	7-10	0.5 – 1	Large wavelets. Crests begin to break; <b>scattered whitecaps.</b>
4	11-16	1 - 2	Small waves with breaking crests. Fairly <b>frequent whitecaps.</b>
5	17-21	2-3	Moderate waves of some length. <b>Many whitecaps.</b> Small amounts of spray.

The team leader will make the decision on when a cetacean is suitable for sampling. This will be based on cetacean behaviour, group demographics as well as sea state (Table 1) and other safety considerations. Only cetaceans in healthy condition (assessed by the appearance of the epaxial muscle mass on either side of the animal's dorsal fin, as in Pugliares et al., 2007 and Clegg et al., 2015) are to be sampled; animals appearing emaciated (skinny) or in poor body condition with a body condition score (BCS) of one are not considered suitable (Table 2). Calves will not be sampled and will be differentiated based on size (<2/3 the length of the mother [i.e., ~1.5m for dolphin calves]) and position in relation to the mother. i.e., dependent calves return frequently to the tailstock region of the mother (Mann & Smuts 1999). Weaned calves (i.e., young juveniles) may be considered suitable to sample.

Table 2 Dolphin Body Condition score (Taken from Clegg et al. 2015)

BCS	DESCRIPTION
1	<u>Emaciated.</u> <i>Head:</i> concave depression behind blowhole. Scapula bones visible. <i>Side view:</i> Ribs clearly visible; hollow area under dorsal fin above spine. <i>Dorsal view:</i> concave indent behind skull ('peanut head'), minimal fat cover
3	<u>Average.</u> <i>Head:</i> fat cover around skull. Scapula bones only visible when pectoral fin is manipulated. <i>Side view:</i> ribs not visible; notable layer of fat parallel and adjacent to spine. <i>Dorsal view:</i> smooth transition from head to neck; rounded fat deposition along spine.



When a sighting has been made and the animals are deemed appropriate for sampling, the shooter should prepare the rifle and dart as described above, and the observer prepare the camera. Photo identification can commence prior to biopsying, but once attention has been turned to the sampling event and a dart loaded, the photographer must move to behind the shooter. The vessel should approach the animal(s) at a speed reduced to no wake or be in neutral and position the vessel to within 5-20m of the cetacean as it surfaces, giving the shooter the best opportunity to fire the dart. The photographer should be prepared (but well back from the shooter) to photograph the dorsal fin of the animal that is to be darted at the same time the dart is fired.

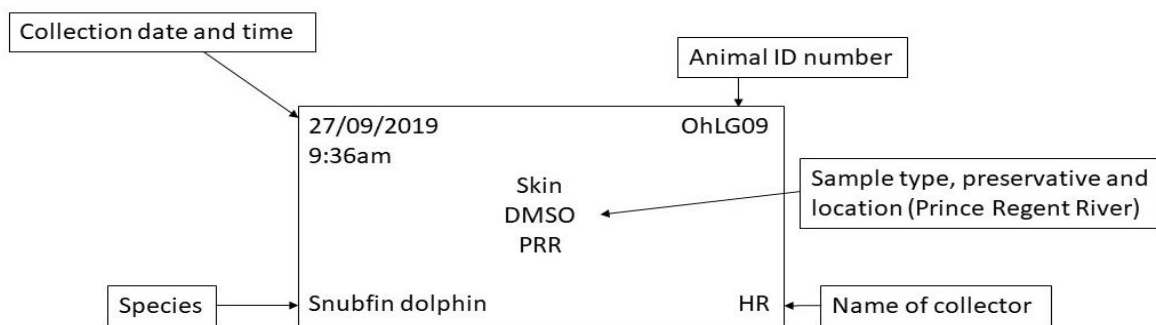
Samples should be taken from in line with and below the dorsal fin, where the blubber layer is thickest. Once fired, the dart will strike the animal, penetrate to a stop (generally 4-5mm into the blubber layer), then bounce off and float at the water/sea surface for recovery. In some whales, the probability of collecting a sample including skin and blubber increases if the angle of impact of the dart is perpendicular to the body surface, but this is less critical when the darts are extremely sharp (Barrett-Lennard, Smith & Ellis 1996). Once the animal(s) is clear of the vessel, the dart is recovered using a hand-net; the sample is extracted using sterile forceps and stored in a small vial containing the appropriate preservative for the purposes of the research question(s) and a label including the sample code (a unique identifier), ID code or name of the individual sampled if known from DolFin catalogue, species name, date,



area/site being sampled (Figure 6 & Figure 7). Appropriate preservatives to be used in the sample vials may include DMSO (Dimethyl sulfoxide a salt saturated solution) or 100% ethanol for genetic molecular analysis), freezing for stable isotope analysis (if possible –80°C freezer if portable one is available) dry ice is a suitable alternative for short periods of time but decays rapidly in warm conditions. A data sheet is completed for every biopsy attempt that includes information on the date, time and location of the sighting; group composition, age class and other information of interest on the group and the behavioural response of the animal(s) to the biopsy attempt (see example Appendix 1, explained further in section 4.5 Data recording). Sterile latex gloves are worn when handling the darts and biopsy heads, to minimize the risk of infection to both the dolphin and observer and cross-contamination by DNA/RNA from handling or other samples. Gloves will be changed between sampling events/animals.

An alternative tool used for sampling is the biopsy pole which is only suited to sampling when cetaceans closely approach the vessel (<1 metre depth or at the surface). Bow-riding cetaceans can be sampled when they are close to the water surface (<1 metre deep) and occasionally when they surface. Sampling will not be attempted on calves (must be >1.5 metres long for dolphins and not swimming in baby position at the tailstock region of the mother). When the team leader deems it suitable, the researcher with the pole should position themselves in the bow of the vessel or on the side closest to the location of the animals. Depending on the depth of the cetacean and the speed of the vessel, the biopsy pole is held and jabbed firmly (but with limited travel range or else the needle may get bent/damaged) into the animal, aiming at the body areas lateral to the base of the dorsal fin. The pole is held at an angle of between 60° and 90° to the water surface, depending on the sampling vessel. A constant speed of 2-6 km/h will be maintained during sampling (Bilgmann et al., 2007).

The biopsy pole is most successful when used on individuals riding the pressure wave at the bow of the vessel. and should only be attempted when the vessel is travelling at slow speed (i.e., a no wake speed). Clear communication must be maintained between the sampler and the vessel master, and the sampler must remain within the vessel, including the bow rail, during the entire sampling procedure. It is a less common sampling method as the opportunities to collect such samples are fewer and dependent on the behaviour and approachability of the target species.



\*Double labelling: inside & outside\*  
 \*Water proof: (use pencil on waterproof paper)\*

Figure 6 Example sample label showing the information to be recorded with each biopsy sample.

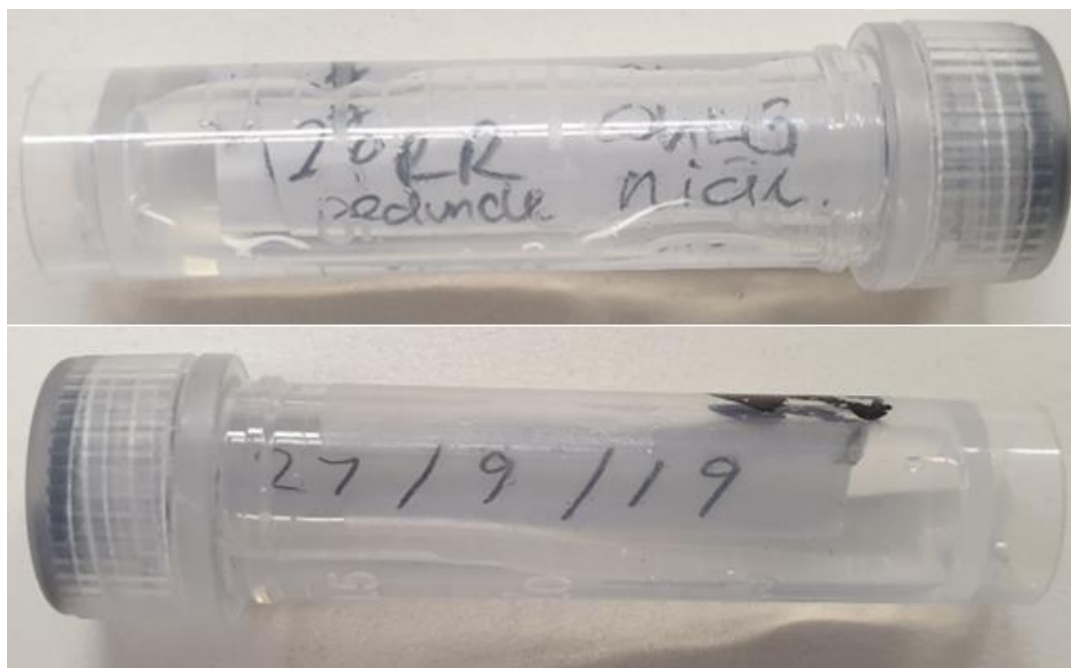


Figure 7 Small vial (approx. 4.5 cm length) containing biopsy sample, appropriate storage solution and label. Label includes sample number taken that day (#2), species (Oh = *Orcaella heinsohni*), area being sampled (LG = Lalang-garram Marine Park, PRR = Prince Regent River), individual's identifying feature (peduncle nick), date sampled (27/9/19).

### 5.5 Data recording

The observer is responsible for filling out the datasheets as the shooter and vessel driver are focussed on the animals. The observer records sighting and survey information, GPS position, sample details, notes on cetacean behavioural response and all other data fields listed in the biopsy sampling datasheet (Appendix 1). GPS coordinates should be taken from the handheld GPS on board with coordinates in decimal degrees and waypoint number noted on the datasheet or if time permits the coordinates transcribed on the datasheet. Familiar dolphins already in the DolFin catalogue with ID codes can be listed in the catalogue ID section (should always be photographed regardless of familiarity to update the catalogue with any changes to the dorsal fin and for the sighting history), as well as sex or age (adult, sub-adult, calf, unknown) of animals if known. Notes on distinctive dorsal fins such as unique scarring, nicks, pigmentation, deformities or 'clean fins' can be drawn or described in comments. If photos are captured of animals, note the frame numbers on the camera using the playback display panel on the camera. The level of response of the sampled animal at the moment of sampling should be recorded (Appendix 1; 0=none, 1=flinch, 2=buck, 3=leap, 4=multiple leaps, O=other) and behaviour afterwards monitored. The observer is responsible for ensuring all data fields are filled in and taking a photo of the datasheet to indicate the end of the sighting. This also serves as a good back up of the datasheet.

At the end of the day, all samples should be stored appropriately for the preservative used i.e., ethanol kept cool and refrigerated if possible but not frozen, DMSO should be kept cool and dry and those samples without preservative should be frozen as soon as possible. All photos from each camera used should be downloaded onto an external hard drive into the trip folder, in a new folder labelled with that day's date (YYYYMMDD). Do not clear SD cards

as they will be used as backups until the photos are transferred on to the server. If operating multiple vessels, the folder should include the vessel name (e.g., YYYYMMDD\_vessel name). Refer to SOP No: TBA Vessel-based Cetacean Surveys Using Photo Identification for detailed instructions for data processing at the end of the day. The datasheets should be reviewed to ensure all information has been recorded on the appropriate datasheets and then kept in a safe location on the mother vessel or accommodation. Once all data is copied onto the external hard drive, make a backup on the second external hard drive. All biopsy sampling equipment (dart heads, probes, forceps etc.) should be sterilised as described above (Section 4.3) and appropriately stored ready for the next day of sampling.

At the end of the field trip most samples can be flown back to Perth as hand luggage. There are no restrictions on transporting samples stored in DMSO (Dimethyl sulfoxide) but a copy of the MSDS is recommended. Biological samples stored in small amounts (no more than 30ml per container and a maximum of 1 L) of flammable liquids such as ethanol can be flown in hand luggage. For frozen samples include ice bricks to keep chilled and prevent defrosting which will compromise DNA quality. Samples that are stored on dry ice must travel as hand luggage and prior permission needs to be sought from the airline before the flight. The dry ice needs to be in an esky that can be vented to allow gas to escape. Samples should be provided to the laboratory for analysis as soon as possible after the field trip to optimise the quality of the DNA.

## 6 Competencies and Approvals

A person who is competent has the knowledge, skills, and experiences that allow them to undertake cetacean biopsy sampling and appropriately manage adverse events as required. Department personnel, and other external parties covered by the department's Animal Ethics Committee, undertaking projects involving biopsy sampling require approval from the committee and will need to satisfy the competency requirements (Table 3). Other groups, organisations or individuals using this SOP to guide their activities are encouraged to also meet these competency requirements as well as their animal welfare legislative obligations.

*Table 3 Competencies for personnel involved in cetacean biopsy sampling*

Competency Category	Competency Requirement	Competency Assessment
<b>Shooter</b>	Minimum current corporate firearm licence (Category E)	Hold a DBCA corporate firearms licence. Trained in category A, B and C3 firearms. Have observed and assisted under the direction of an experienced shooting mentor, until the mentor is satisfied that the mentee is competent in biopsying free-ranging cetaceans. Assessment can be made on target on water prior to sampling.
<b>Photographer / Observer</b>	Prior experience – competency requirement.	Have observed and assisted under the direction of an experienced supervisor as a photographer, until the supervisor is satisfied that a required level of expertise has been achieved.

<b>Vessel operator</b>	Minimum of Coxswain 2 Near Coastal certificate.	AMSA approved assessment. Note there may be limitations to the area of operation.
<b>Secondary observer / Data recorder</b>	Prior training, experience ideal – competency requirement.	Have observed and assisted under the direction of an experienced supervisor, until the supervisor is satisfied that a required level of expertise has been achieved.

In conjunction with possessing the required understanding and knowledge of sampling cetacean using a remote biopsy system and animal welfare requirements, a guide to the experience and skill requirements for personnel to be considered competent to biopsy sampling is as follows: (noting that some personnel with experience may still require initial supervision in unfamiliar locations or with species that they have not encountered previously):

- Recency of time in field: within the past 10 years.
- Minimum 2 individuals of a variety of species handled.

### 6.1 Occupational Health and Safety

Extreme care should be taken when using firearms on a vessel. When attempting to biopsy sample, ensure a job safety analysis has been approved for the work and all personnel involved are familiar with it. Follow the safe rifle procedures for working on a vessel outlined above (Section 4.3). Ensure the layout of the chosen working vessel and positioning of personnel in the vessel allows for clear communication between shooter and vessel master at all times (i.e., centre console windscreen is not obstructing communication). It is a requirement that an experienced cetacean researcher/sampler is always present to assess the level of risk of each sample attempt.

A first aid kit, satellite phone and VHF radio should be carried in the vessel at all times. You must be aware of your own safety and the safety of others. Extreme care should be taken when working in tidal estuaries to avoid exposure to disease carrying insects, like mosquitos and sand-flies. All injuries (even superficial ones) and bites should be appropriately treated as soon as possible to prevent infection and promote healing.

Departmental personnel, contractors and volunteers have duties and responsibilities under the Occupational Safety and Health Act 1984 and Occupational Safety and Health Regulations 1996 to ensure the health and safety of all involved. Fieldwork is to be undertaken in line with the department's corporate guidelines, policies and standard operating procedures, including but not limited to, risk management and job safety analyses. Further information can be found at

<https://dpaw.sharepoint.com/Divisions/corporate/people-services/HS/SitePages/SOPs.aspx>

If department personnel or volunteers are injured, please refer to the departmental Health, Safety and Wellbeing Section's 'Reporting Hazards, Near-misses and Incidents' intranet page, which can be found at <https://dpaw.sharepoint.com/Divisions/corporate/people-services/HS/SitePages/Reporting-Hazards,-Near-Misses-and-Incidents.aspx>

### 6.2 Personal Protective Equipment

Protective gear required shall be determined by the project specific JSA. This may include but

is not limited to adequate sun protection (i.e., long-sleeved clothing, sunscreen, hat, face buff, hand gloves, boat shoes, polarised sunglasses), protection from wind chill and other changes in weather (raincoat may be required), latex gloves.

### 6.3 Manual handling and working with ropes

Manual handling and working with ropes can be an integral part of working with small boats with detachable outboards and there is a high risk of injury (muscular or back). Further advice is available to DBCA personnel by accessing Departmental guidance notes on manual handling.

### 6.4 Working around the water's edge and from vessels

PDF type 1 Life jackets must be worn by those working on vessels. Refer to Department of Parks and Wildlife Corporate Policy Statement No. 84 Boating.

Care must be exercised to avoid slipping into the water from the vessel or when working on riverbanks and boat ramps. All limbs must be kept within the vessel at all times.

## 7 Further Reading

The following documents have been mentioned in the advice regarding cetacean biopsy surveys. It is recommended these documents and others listed below are also considered when proposing to undertake remote biopsy sampling.

- SOP: *Vessel-based Cetacean Surveys Using Photo Identification*.
- Noren, D.P. and Mocklin, J.A. (2012), Review of cetacean biopsy techniques: Factors contributing to successful sample collection and physiological and behavioural impacts. *Marine Mammal Science*, 28: 154-199.
- Raudino, H.C., D'Cruz, E.C., Barrow, D., Waples, K.A. (2019). Dambimangari Rangers Joint Management tropical inshore dolphin survey in Lalang-garram/Camden Sound Marine Park, 23 -30 Sept 2019. Department of Biodiversity, Conservation and Attractions, Perth, Western Australia.
- Raudino, H.C., Douglas, C.R., D'Cruz, E.C., Waples, K.A. (2019). Snubfin dolphin census in Yawuru Nagulagan/Roebuck Bay. Department of Biodiversity, Conservation and Attractions, Perth, Western Australia.
- Raudino, H. C., Tyne, J. A., Smith, A., Ottewell, K., McArthur, S., Kopps, A. M., Chabanne, D., Harcourt, R. G., Pirotta, V., and Waples, K. 2019. Challenges of collecting blow from small cetaceans. *Ecosphere* 10(10):e02901. 10.1002/ecs2.2901

Department of Parks and Wildlife Corporate Policy Statement No. 84 Boating

Department of Parks and Wildlife Boating Code of Practice 2016

*Firearms Act 1973*

*Firearms Regulations 1974*

*Occupational Safety and Health Act 1984*

*Occupational Safety and Health Regulations 1996*

*Biodiversity Conservation Act 2016*

*Biodiversity Conservation Regulations 2018.*

## 8 References

Barrett-Lennard, L., Smith, T.G., Ellis, G.M., 1996. A cetacean biopsy system using lightweight pneumatic darts, and its effect on the behaviour of killer whales. *Marine Mammal Science*, 12, pp. 14-27.

Bearzi, G. 2000. First report of a common dolphin (*Delphinus delphis*) death following penetration of a biopsy dart. *Journal of Cetacean Research and Management* 2: 217– 221.

Bilgmann, K., Griffiths, O.J., Allen, S.J., Moller, L.M., 2007. A biopsy pole system for bow-riding dolphins: sampling success, behavioral responses, and test for sampling bias. *Marine Mammal Science* 23:218–225

Corkeron, P.J., Morris, R.J., Bryden, M.M., 1987. A note on healing on large wounds in bottlenose dolphins, *Tursiops truncatus*. *Aquatic Mammals* 13, 96-98.

Gimenez, J., De Stephanis, R., Gauffier, P., Esteban, R. & Verborgh, P., 2011. Biopsy wound healing in long-finned pilot whales (*Globicephala melas*). *The Veterinary record*, 168, 101.

Krützen, M., Barré, L.M., Möller, L.M., Heithaus, M.R., Simms, C., Sherwin, W.B., 2002. A biopsy system for small cetaceans: darting success and wound healing in *Tursiops* spp. *Marine Mammal Science* 18, 863–878.

Mann, J., and Smuts, B., 1999. Behavioral development in wild bottlenose dolphin newborns (*Tursiops* sp.). *Behaviour* 136(5), 529-566.

Mijele, D., Omondi, P., Gakuya, F., Rossi, L., Chiyo, P.I., Soriguer, R.C., Angelone-Alasaad, S., 2016. A practical guideline to remote biopsy darting of wildebeests for genetic sampling. *International Journal of Veterinary Science and Medicine*. 4. 27-32.

Orams, M.B., Deakin, R.B., 1997. Report on the healing of a large wound in a Bottlenose Dolphin *Tursiops truncatus*. In: M. Hindell, C.M. Kemper (Eds.), *Marine Mammal Research in the Southern Hemisphere Volume I: Status, Ecology and Medicine*. Beatty and Sons, Chipping Norton, pp. 170-173.

Pugliares, K.R., Bogomolni, A., Touhey, K.M., Herzig, S.M., Harry, C.T., Moore, M.J., 2007. *Marine mammal necropsy: an introductory guide for stranding responders and field biologists*. Woods Hole Oceanographic Institution Technical Report WHOI-2007-06, p. 55.

Tezanos-Pinto, G. & Baker, C. S., 2012. Short-term reactions and long-term responses of bottlenose dolphins (*Tursiops truncatus*) to remote biopsy sampling. *New Zealand Journal of Marine and Freshwater Research*, 46, 13-29.

Weller, D. W., Cockcroft, V. G., Würsig, B., Lynn, S. K. & Fertl, D., 1997. Behavioral responses of bottlenose dolphins to remote biopsy sampling and observations of surgical biopsy wound healing. *Aquatic Mammals*, 23, 49-58.

## 9 Glossary of Terms

**Cetacean:** Marine mammals commonly known as whales, dolphins and porpoises.

**Tissue:** groups of cells that have similar structure with a specific function, in a multicellular organism. For the purposes of this SOP, tissue will most often refer to the skin and underlying blubber layer of a cetacean.

**Sample:** a small piece of tissue, intended as a representative of the whole

**DNA (deoxyribonucleic acid):** a macromolecule found in all living cells that contains genetic identifying information of organisms

**Dorsal fin:** the fin located on the top side of an animal, commonly used to identify species.

**Beaufort Sea State (BSS):** scale for estimating wind strengths based on appearance of the sea surface. Often used by researchers as it affects the probability of sighting cetaceans.